

DE-O-METHYLDIAPORTHIN, A PHYTOTOXIN FROM *DRECHSLERA SICCANS*

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Key Word Index—*Drechslera siccans*; isocoumarin; phytotoxin; diaporthin; de-*O*-methyldiaporthin.

Abstract—A phytotoxic metabolite has been isolated from cultures of *Drechslera siccans* and structurally characterized as 6,8-dihydroxy-3-(2'-hydroxypropyl) isocoumarin (de-*O*-methyldiaporthin).

INTRODUCTION

Drechslera siccans is a pathogenic fungus on oats (*Avena sativa* 'Park'), perennial ryegrass (*Lolium perenne* L.) and Italian ryegrass (*Lolium multiflorum* Lam.). Fungal infection causes irregular dark brown to reddish brown spots (0.5–1 mm long and 0.2–0.5 mm wide) on the leaves of these hosts within one week of inoculation [1]. Occasionally, these lesions have tan centres and assume an eyespot appearance. Our interest in new phytotoxic compounds from the genus *Drechslera* led us to study *Drechslera siccans*, and we now report the isolation and structural elucidation of a new phytotoxic isocoumarin, de-*O*-methyldiaporthin.

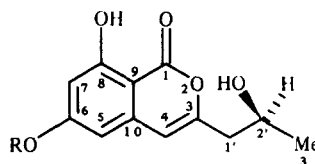
RESULTS AND DISCUSSION

Isolates of *D. siccans* were cultured in M-1-D media using our previously reported procedure [2, 3]. Culture

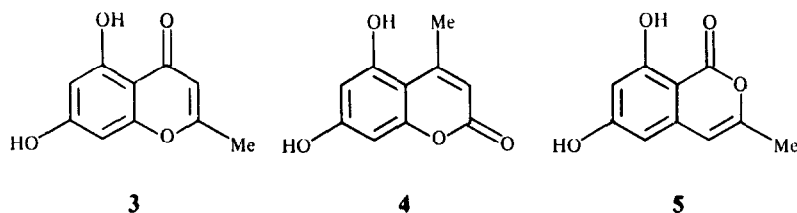
filtrates were extracted with ethyl acetate and the title compound was isolated by successive preparative TLC and further purified by reverse phase HPLC to yield pure **1** as a white solid.

The high resolution mass spectra of **1** gave $[M]^+$ at m/z 236.0690, corresponding to the molecular formula $C_{12}H_{12}O_5$ (236.0685). The UV spectra (Table 1) indicated a conjugated aromatic chromophore [4]. The 1H NMR also indicated that an aromatic ring was part of the structure. Two doublets at δ 6.38 and 6.42 with a small coupling constant, $J = 2.17$ Hz, suggested two aromatic protons with a *meta* relationship. In addition, the 1H NMR showed signals corresponding to a methyl group (δ 1.24), a methylene group (δ 2.60) and an olefinic proton (δ 6.44). A signal at δ 11.18 was interpreted as a hydrogen bonded phenolic proton.

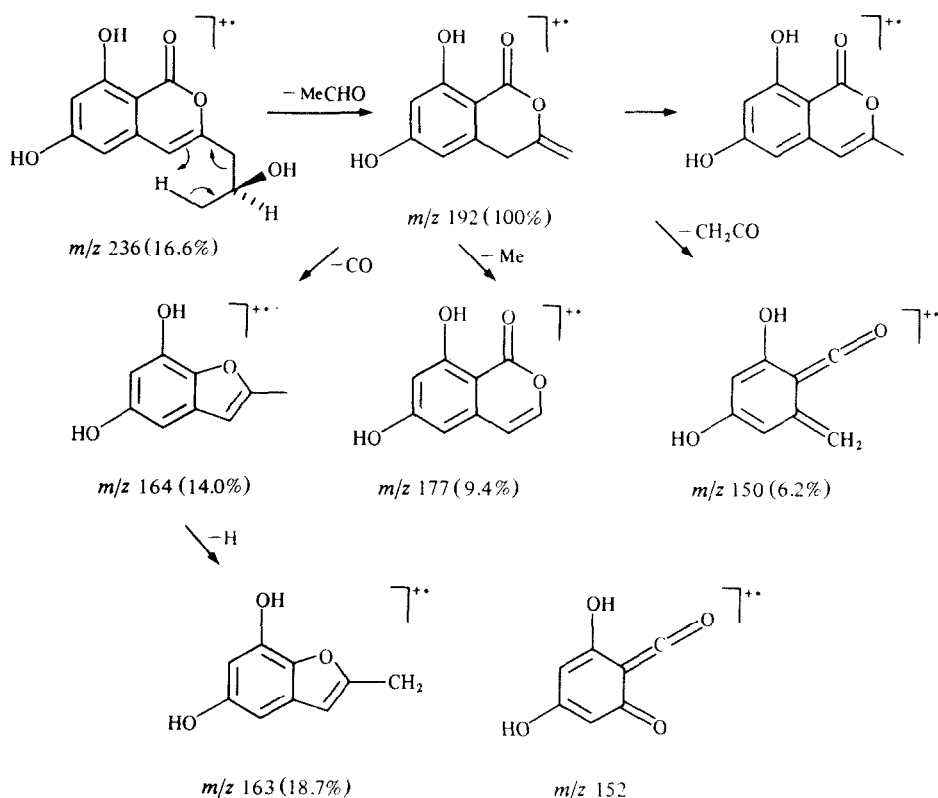
The mass spectral fragmentation pattern revealed a benzopyran as the major molecular building block. Loss of 44 mass units to give the high intensity peak at m/z 192 in the mass spectra was consistent with the benzopyran structure, leaving the methyl and methylene groups as part of the side chain. Lack of a peak at m/z 152 ruled out a chromone structure, since this peak is typically ob-



- 1** R = H de-*O*-methyldiaporthin
2 R = Me diaporthin



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served for γ -benzopyrans [5] (Scheme 1, bottom right). The most plausible structure for **1** appeared to be an 6,8-dihydroxy-4-methylcoumarin (**4**) [7] and 6,8-dihydroxy-3-methylisocoumarin (**5**) [8], shown in Table 1. It was further confirmed by comparing spectroscopic data with the known compound diaporthin (**2**), a fungal metabolite from *Endothia parasitica* [9].

This identification was supported by an examination of the UV data of 5,7-dihydroxy-2-methylchromone (**3**) [6], 5,7-dihydroxy-4-methylcoumarin (**4**) [7] and 6,8-dihydroxy-3-methylisocoumarin (**5**) [8], shown in Table 1. It was further confirmed by comparing spectroscopic data with the known compound diaporthin (**2**), a fungal metabolite from *Endothia parasitica* [9].

Bioassay data are presented in Table 2. Host plants of this pathogen showed little or no sensitivity to de-O-methyldiaporthin (**1**) (Table 2). However, corn, crabgrass, and soybean reacted to 1 μ g (4 nmol). Barnyard grass and

spiny amaranth were sensitive to 8 and 21 nmol, respectively.

EXPERIMENTAL

Fungal isolates and cultures. Isolates of *D. siccans* were kindly provided by Dr Richard Morrison at Northrup King Co., Woodland, California. Cultures were maintained on potato dextrose agar with 18% V-8 juice (v/v). Broth cultures were prepared in M-1-D medium and fermented for 21 days at 25° in a shaker (200 rpm).

Isolation of de-O-Methyldiaporthin (1). The fungal culture was filtered through four layers of cheese cloth. The filtrate was extracted with EtOAc (3 \times 1/3 vol) and concd *in vacuo*. The residue was taken up in MeOH, applied to a Sephadex LH-20 column and eluted with MeOH. The major fraction was then separated by prep TLC (20 \times 20 cm, 1000 μ m) using CHCl₃-MeOH (9:1). The fraction with an *R_f* of 0.38 was prep. TLCed again and further purified by reverse phase HPLC (4.6 \times 250 mm Supelco column, UV detector at 254 nm, MeCN-H₂O, linear gradient program, 0-100% MeCN, 1 ml/min, *R_t* 18.4 min). Evaporation from Me₂CO gave de-O-methyldiaporthin (**1**) as a white solid. The yield was ca 3 mg/l of culture.

De-O-methyldiaporthin HREIMS: C₁₂H₁₂O₅, obsd 236.0690, calcd 236.0685. LREIMS (probe) 70 eV, *m/z* (rel. int.): 236 (17), 221 (3), 193 (15), 192 (100), 177 (9), 174 (8), 164 (14), 163 (19), 150 (6), 146 (24), 135 (4), 121 (21). [α]_D + 22° (MeOH, c 0.09). UV (see Table 1). ¹H NMR (400 MHz, acetone-*d*₆): δ 1.24 (d, 3H, *J* = 6.2 Hz), 2.60 (m, 2H), 4.17 (m, 1H), 6.38 (d, 1H, *J* = 2.17 Hz), 6.42 (d, 1H, *J* = 2.17 Hz), 6.44 (s, 1H), 11.18 (s, 1H). ¹³C NMR: δ 23.4 (q,

Table 1. UV data of compounds **1**, **3**-**5**

1	$\lambda_{\text{max}}^{\text{MeOH}}$ (nm) (log ϵ)	3	4	5
237 (4.31)	227 (4.08)	—	—	237 (4.62)
246 (4.38)	249 (4.13)	—	—	245 (4.69)
275 (3.68)	256 (4.13)	258 (3.82)	—	278 (3.85)
289 (3.53)	295 (3.65)	—	—	289 (3.72)
326 (3.52)	—	322 (4.10)	—	324 (3.79)

Table 2. Sensitivity of various plants to de-O-methyldiaporthin (1)

Plant		Necrotic area (square mm)			
		10 µg	5 µg	2 µg	1 µg
Perennial rye	<i>Lolium perenne</i> L.	0	0	0	0
Park oat	<i>Avena sativa</i> L.	1	<1	0	0
Corn (W64A-N)	<i>Zea mays</i> L.	—	24	15	4
Crabgrass	<i>Digitaria ischaemum</i> Schreb.	12	16	6	1
Barnyard grass	<i>Echinochloa crus-galli</i> L.	12	9	2	0
Yellow nutsedge	<i>Cyperus esculentus</i> L.	0	0	0	0
Soybean	<i>Glycine max</i> L.	12	9	9	2
Rape (Harosoy 63)	<i>Brassica campestris</i> L.	0	0	0	0
Spiny amaranth	<i>Amaranthus spinosus</i> L.	3	1	0	0

Specified amounts were applied in 5% aqueous ethanol (3 µl). Necrotic area was measured after 48 hr.

C3'), 44.1 (t, C1'), 65.7 (d, C2'), 99.2 (d), 102.4 (d), 103.8 (s), 106.6 (s), 140.5 (d, C4), 156.3 (s, C3), 164.5 (s), 167.2 (s), 167.8 (s).

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